

Complete Genome Sequence of *Arthrobacter globiformis* Bacteriophage Ren19

Lauren A. Holt¹, Jazmine N. Taylor², Kera S. Ridley³, Devin-Danielle V. Webb³, David S.I. Davenport¹, He Fu^{4§}

¹Department of Chemical, Biological and Bioengineering, North Carolina A&T State University, Greensboro, NC, United States

²Department of Physics, North Carolina A&T State University, Greensboro, NC, United States

³Department of Animal Sciences, North Carolina A&T State University, Greensboro, NC, United States

⁴Department of Biology, North Carolina A&T State University, Greensboro, NC, United States

§To whom correspondence should be addressed: hfu@ncat.edu

Abstract

Bacteriophage Ren19, isolated from a soil sample collected in Greensboro, North Carolina, USA, exhibits a siphovirus morphology and infects *Arthrobacter globiformis* B-2979. Its 68,603-bp genome has a GC content of 65.4% and contains 118 predicted genes. Based on gene-content similarity, Ren19 is assigned to actinobacteriophage cluster AP2.

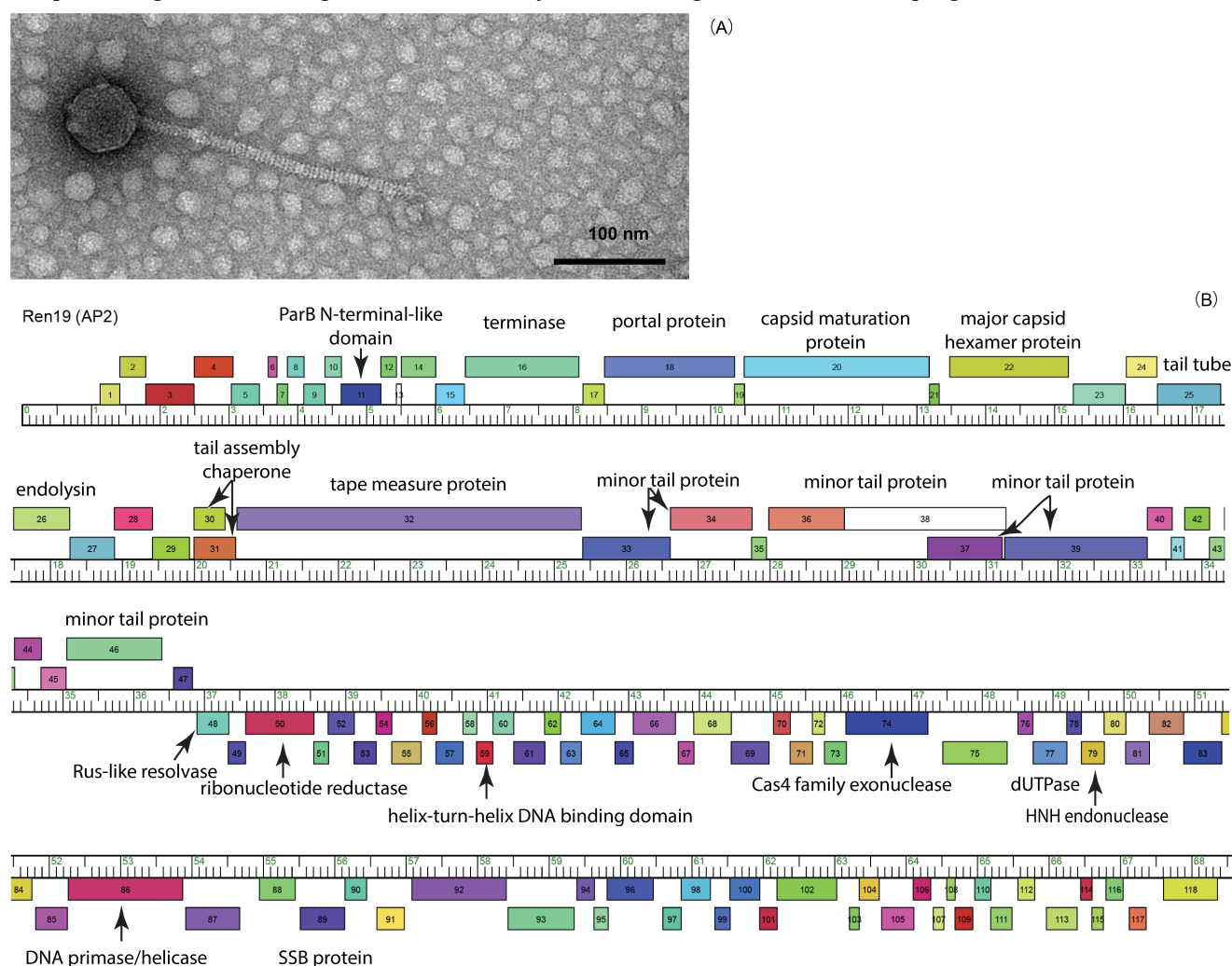


Figure 1. Transmission electron micrograph and genome map of bacteriophage Ren19:

(A) Negative-stain transmission electron micrograph of Ren19 stained with 1% uranyl acetate. Ren19 has a capsid diameter of 58 nm and a tail length of 241 nm. Imaging was performed at the University of Maryland, Baltimore County. (B) Genome map of Ren19. Predicted genes are represented by boxes, with the corresponding gene numbers shown inside. Genomic coordinates are indicated by the ruler, with each tick mark representing 100 bp. The map was generated using Phamerator and the Actino_Draft_655 database (Cresawn et al., 2011).

Description

Bacteriophages are viruses that infect bacteria and constitute one of the most abundant and diverse biological entities on Earth (Hatfull, 2022). Their ability to selectively infect and kill bacterial hosts has generated growing interest in their therapeutic use against antibiotic-resistant infections (Sawa et al., 2024). Because most phages have relatively narrow host ranges, the continued isolation and characterization of novel bacteriophages are essential for expanding the repertoire of potential therapeutic candidates. To this end, we report the isolation and characterization of Ren19, a novel bacteriophage recovered from flowerbed soil on the campus of North Carolina Agricultural and Technical State University in Greensboro, North Carolina, USA (36.079194° N, 79.772028° W).

Ren19 was isolated using a modified direct-isolation procedure (Zorawik et al., 2024). Briefly, the soil sample was suspended in phage buffer by vigorous shaking for 3 h, allowed to settle overnight at 4°C, and filtered through a 0.22-µm pore-size filter. The filtrate was plated with *A. globiformis* B-2979 in PYCa top agar and incubated at 30°C for 48 h. Ren19 produced small, clear plaques (1.4 mm diameter, *n* = 5) and was purified through two successive rounds of plaque isolation. Negative-stain transmission electron microscopy using 1% uranyl acetate revealed a siphovirus morphology, with a capsid diameter of 58 nm and a tail length of 241 nm (*n* = 1; Fig. 1A). Dimensions were measured using ImageJ version 1.54 (Schneider et al., 2012).

A lysate for Ren19 was prepared (1.5 × 10¹⁰ PFU/mL) and used to extract DNA with the Promega Wizard DNA kit. Phage DNA was then prepared for sequencing with the NEB Ultra FS kit and sequenced on an Illumina NextSeq 1000 (XLEAP-P1 kit), yielding 506k single-end 100 base reads. Raw sequencing reads were trimmed using cutadapt v4.7 (using the option: `-nextseq-trim 30`) and subsequently filtered with skewer v0.2.2 (using the options: `-q 20 -Q 30 -n -l 50`) prior to assembly. The genome was assembled using Newbler v.29 (Miller et al., 2010) and checked for completeness using Consed v2.9 (Gordon et al., 1998), resulting in an assembled genome of 68,603 base pairs with 683-fold coverage. The genome consisted of 65.4 % GC content, with a direct terminal repeat of 614 base pairs.

Glimmer v. 3.02 (Delcher et al., 1999), and Genemark v. 2.5p (Besemer & Borodovsky, 2005) were used to auto-annotate the bacteriophage genome in DNA Master (Pope & Jacobs-Sera, 2018). The auto-annotation was refined using Genemark, Phamerator (using Actino_draft database v655) (Cresawn et al., 2011), Starterator (phages.wustl.edu/starterator/), and BLAST (Sayers et al., 2025) using the Actinobacteriophage and NCBI non-redundant databases. Gene function was assigned in PECAAN (<https://discover.kbrinsgd.org>) and refined in PHEONA (<https://www.pheona.org/>) using HHPred searches against the PDB_mmCIF70, Pfam- v.36, NCBI Conserved Domains databases (Söding et al., 2005) and transmembrane domain containing proteins predicted with DeepTMHMM v. 1.0.42 (Hallgren et al., 2022). No tRNAs were found for this phage using ARAGORN (<https://www.trna.se/>). Default parameters were used for all software.

Based on a gene-content similarity of at least 35% to phages in the Actinobacteriophage Database (PhagesDB), Ren19 was assigned to cluster AP2 (Pope et al., 2017; Russell and Hatfull, 2016). Consistent with the genomic organization of other cluster AP2 siphoviruses, the first 47 genes are transcribed in the forward direction, whereas the remaining genes are transcribed in the reverse direction. Putative functions were assigned to 34 of the 118 predicted genes (Figure 1B). Ren19 encodes many proteins commonly found in bacteriophages, including a terminase, portal protein, major capsid protein, major tail protein, tape-measure protein, tail-tube protein, and minor tail protein. The genome also contains identifiable genes encoding an endolysin, HNH endonuclease, tail-assembly chaperones, and capsid-maturation protease. Two enzymes involved in nucleic acid replication or recombination—a DNA primase/helicase and a RusA-like resolvase—were also identified. Like other characterized cluster AP2 phages, Ren19 lacks identifiable integrase and repressor genes, suggesting that it is unlikely to establish lysogeny.

Data availability.

Annotated genome sequence can be accessed for Ren19 at GenBank with Accession Number PZ744772. Sequence reads are deposited at NCBI under SRA accession number [SRX31950264](https://www.ncbi.nlm.nih.gov/sra/SRX31950264).

Acknowledgements: We would like to thank the Howard Hughes Medical Institute for support of the SEA-PHAGES program.

References

- Besemer J, Borodovsky M. 2005. GeneMark: web software for gene finding in prokaryotes, eukaryotes and viruses. *Nucleic Acids Research* 33: W451-W454. DOI: [10.1093/nar/gki487](https://doi.org/10.1093/nar/gki487)
- Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. *BMC Bioinformatics* 12: 10.1186/1471-2105-12-395. DOI: [10.1186/1471-2105-12-395](https://doi.org/10.1186/1471-2105-12-395)
- Delcher A. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Research* 27: 4636-4641. DOI: [10.1093/nar/27.23.4636](https://doi.org/10.1093/nar/27.23.4636)

Gordon D, Abajian C, Green P. 1998. *Consed*: A Graphical Tool for Sequence Finishing. *Genome Research* 8: 195-202. DOI: [10.1101/gr.8.3.195](https://doi.org/10.1101/gr.8.3.195)

Hallgren J, Tsirigos KD, Pedersen MD, Almagro Armenteros JJ, Marcatili P, Nielsen H, Krogh A, Winther O. 2022. DeepTMHMM predicts alpha and beta transmembrane proteins using deep neural networks. : 10.1101/2022.04.08.487609. DOI: [10.1101/2022.04.08.487609](https://doi.org/10.1101/2022.04.08.487609)

Hatfull GF. 2022. Mycobacteriophages: From Petri dish to patient. *PLOS Pathogens* 18: e1010602. DOI: [10.1371/journal.ppat.1010602](https://doi.org/10.1371/journal.ppat.1010602)

Pope WH, Mavrich TN, Garlena RA, Guerrero-Bustamante CA, Jacobs-Sera D, Montgomery MT, et al., Hatfull. 2017. Bacteriophages of *Gordonia* spp. Display a Spectrum of Diversity and Genetic Relationships. *mBio* 8: 10.1128/mbio.01069-17. DOI: [10.1128/mbio.01069-17](https://doi.org/10.1128/mbio.01069-17)

Pope WH, Jacobs-Sera D. 2017. Annotation of Bacteriophage Genome Sequences Using DNA Master: An Overview. *Methods in Molecular Biology, Bacteriophages* : 217-229. DOI: [10.1007/978-1-4939-7343-9_16](https://doi.org/10.1007/978-1-4939-7343-9_16)

Russell DA, Hatfull GF. 2016. PhagesDB: the actinobacteriophage database. *Bioinformatics* 33: 784-786. DOI: [10.1093/bioinformatics/btw711](https://doi.org/10.1093/bioinformatics/btw711)

Russell DA. 2017. Sequencing, Assembling, and Finishing Complete Bacteriophage Genomes. *Methods in Molecular Biology, Bacteriophages* : 109-125. DOI: [10.1007/978-1-4939-7343-9_9](https://doi.org/10.1007/978-1-4939-7343-9_9)

Sawa T, Moriyama K, Kinoshita M. 2024. Current status of bacteriophage therapy for severe bacterial infections. *Journal of Intensive Care* 12: 10.1186/s40560-024-00759-7. DOI: [10.1186/s40560-024-00759-7](https://doi.org/10.1186/s40560-024-00759-7)

Sayers EW, Beck J, Bolton EE, Brister JR, Chan J, Connor R, et al., Pruitt. 2024. Database resources of the National Center for Biotechnology Information in 2025. *Nucleic Acids Research* 53: D20-D29. DOI: [10.1093/nar/gkae979](https://doi.org/10.1093/nar/gkae979)

Schneider CA, Rasband WS, Eliceiri KW. 2012. NIH Image to ImageJ: 25 years of image analysis. *Nature Methods* 9: 671-675. DOI: [10.1038/nmeth.2089](https://doi.org/10.1038/nmeth.2089)

Soding J, Biegert A, Lupas AN. 2005. The HHpred interactive server for protein homology detection and structure prediction. *Nucleic Acids Research* 33: W244-W248. DOI: [10.1093/nar/gki408](https://doi.org/10.1093/nar/gki408)

Zorawik M, Jacobs-Sera D, Freise AC, SEA-PHAGES, Reddi K. 2024. Isolation of Bacteriophages on Actinobacteria Hosts. *Methods in Molecular Biology, Phage Engineering and Analysis* : 273-298. DOI: [10.1007/978-1-0716-3798-2_17](https://doi.org/10.1007/978-1-0716-3798-2_17)

Funding: LAH and JNT were supported by the Expanding Undergraduate Research Engagement in the Sciences initiative through the Building Success in Science (SiS) Learning Community. HF was supported by funding from the Howard Hughes Medical Institute to expand the SEA-PHAGES program at North Carolina Agricultural and Technical State University.

Conflicts of Interest: The authors declare that there are no conflicts of interest present.

Author Contributions: Lauren A. Holt: investigation, formal analysis. Jazmine N. Taylor: investigation, formal analysis. Kera S. Ridley: investigation, formal analysis. Devin-Danielle V. Webb: investigation, formal analysis. David S.I. Davenport: formal analysis, investigation. He Fu: conceptualization, writing - original draft, writing - review editing, supervision.

Reviewed By: Anonymous

History: Received August 17, 2026 **Revision Received** September 18, 2026 **Accepted** September 25, 2026 **Published Online** September 28, 2026 **Indexed** October 12, 2026

Copyright: © 2026 by the authors. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC BY 4.0) License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Citation: Holt LA, Taylor JN, Ridley KS, Webb DDV, Davenport DSI, Fu H. 2026. Complete Genome Sequence of *Arthrobacter globiformis* Bacteriophage Ren19. *microPublication Biology*. [10.17912/micropub.biology.002350](https://doi.org/10.17912/micropub.biology.002350)